Grape-berry moth - Eupoecilia ambiguella Hbn.

The body of the moth is 6-7 mm long, the wingspan is 13-15 mm. The forewings are pale yellow to straw-yellow, with a characteristic, trapezoid brown or black stripe, a "saddle". The hindwings are greyish white, with brownish grey edges. The host plants of the larvae include grapes, currants. Larvae of the first larval generation in the spring damage the flowers, especially those which are near to the soil level. The second larval generation damages the grape berries, forming a tube-like webbing on the bunches.

The CSALOMON® pheromone trap should be suspended inside the vineyards, preferably on leafy shoots at the height of the bunches of berries (depending on the mode of culture; usually at 1.0-1.5 m). Usual beginning of trapping in Hungary is middle of April.

Selectivity of the CSALOMON® trap (based on tests performed in Hungary): in the vicinity of forests the trap can capture sizeable numbers of *Cnephasia* spp., first of all *C. ecullyana*, which is somewhat smaller than the grape-berry moth, and its forewings are uniformly dark greyish black.

Longevity of the CSALOMON® trap in field conditions: depending on the warmth

trap for most effective detection and monitoring. Renewal of sticky inserts in intervals of 7-10 days. In case of high catches this may become necessary more often. Pheromone trapping is a very widespread and an almost exclusive

of the weather at least 4-6 weeks. After this period we suggest to set up a new

method all over Europe to detect the occurrence and to monitor the flight pattern



The larva and its damage, which should be averted





of this pest. For best results insecticide treatments should be timed after 10 -14 daysfollowing the peak flight; according to experience in Hungary it is usually not necessary to spray if the weekly catch does not exceed 25-30 moths per trap[1]. Please note that these thresholds may change according to geographic area, climatic conditions, etc.

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The moth

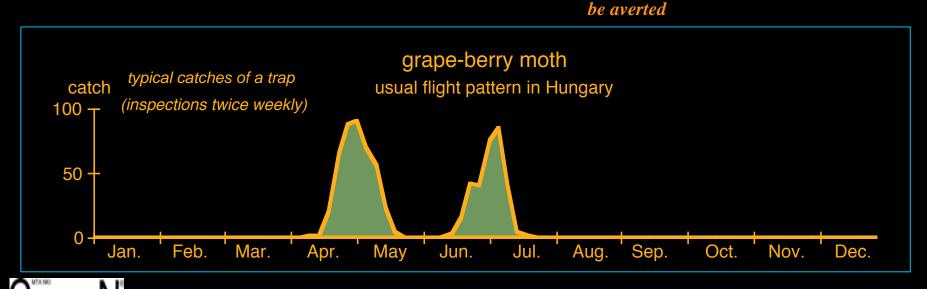
which is

captured in the

Results and methods of monitoring in other countries are given by several authors[2].

[1] Diófási and Sélley: Agrofórum 6:29, 1995. [2] Vogt H. Journal Of Applied Entomology-zeitschrift Fur Angewandte Entomologie 115:217-232, 1993 Rauscher S. J. Chem. Ecol. 10:253-264, 1984 Arn H. J. Chem. Ecol. 12:1417-1429, 1986 Arn H. Mitt. Schweiz. Entomol. Ges. 52:49-55, 1979 Arn H. Mitt. Schweiz. Entomol. Ges. 52, 45-48, 1979 Rauscher S. Entomol. Exp. Appl. 25:16-20, 1979 Roehrich R. Vititechnique. 31:3, 1980 Rauscher S. J. Chem. Ecol. 10:253-264, 1984





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So it looks when caught in the CSALOMON® RAG trap!